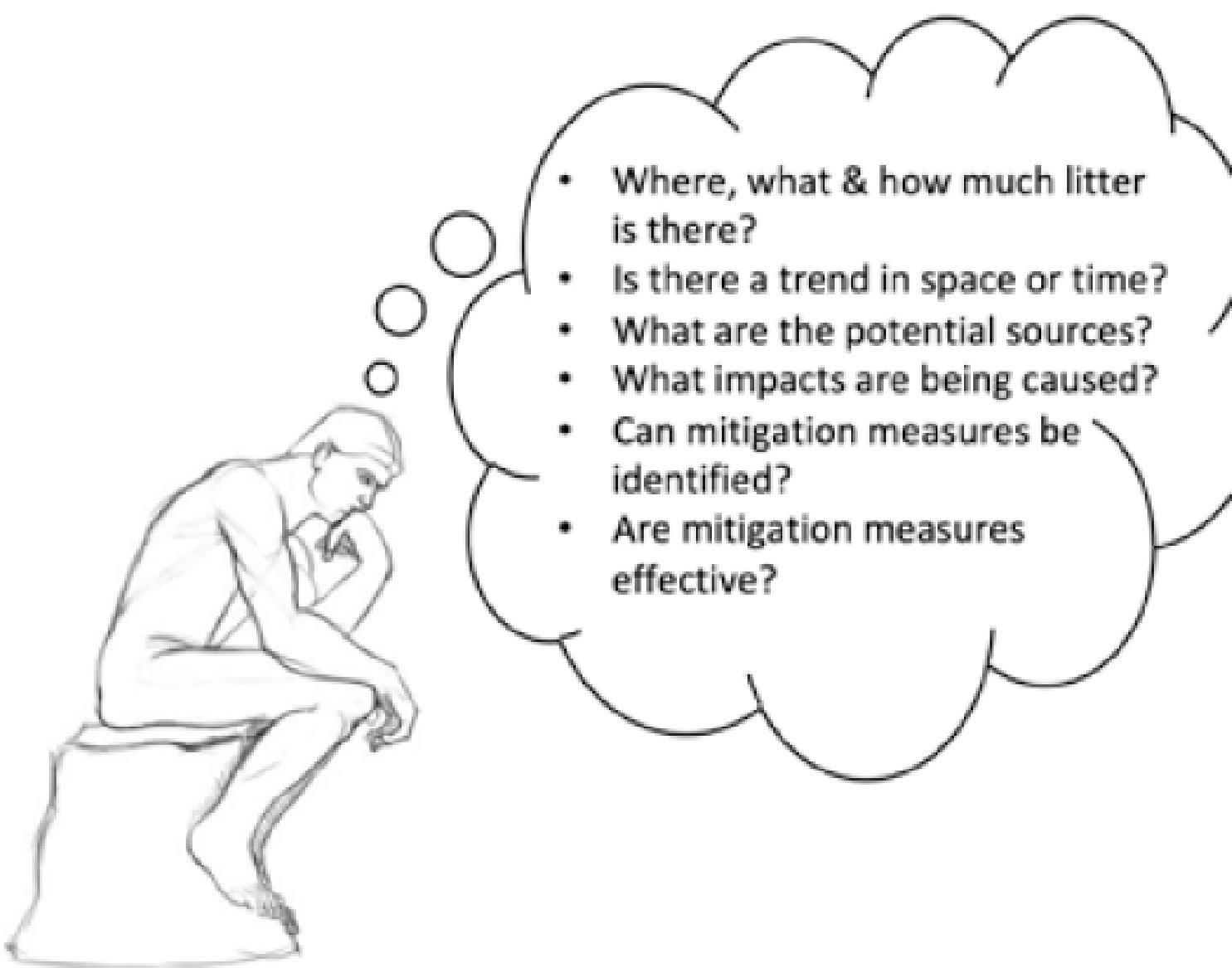




GUIDELINES FOR THE MONITORING AND ASSESSMENT OF PLASTIC LITTER IN THE OCEAN



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What do you want to know?

- Where, what & how much litter is there?
- Is there a trend in space or time?
- What are the potential sources?
- What impacts are being caused?
- Can mitigation measures be identified?
- Are mitigation measures effective?

Where do you look?

SHORELINE

SEA
SURFACE &
WATER
COLUMN

SEA FLOOR

BIOTA

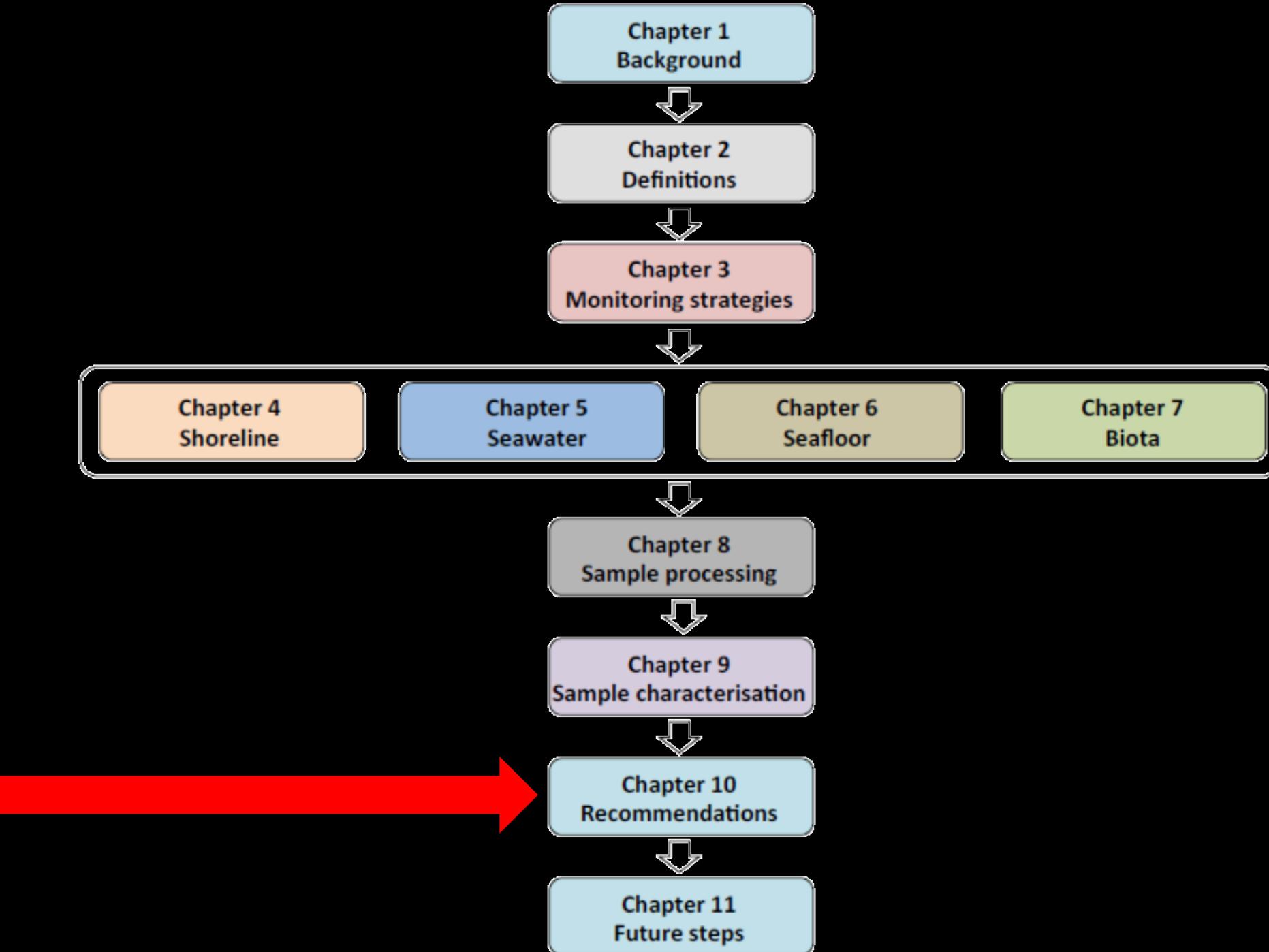


Table 10.3: Estimated costs and level of expertise for the different protocols adapted from Table 10.2. L: Low (< 10K USD); M Medium (<50K USD); H High (<100K USD); VH Very High (>100K USD). Vehicles.

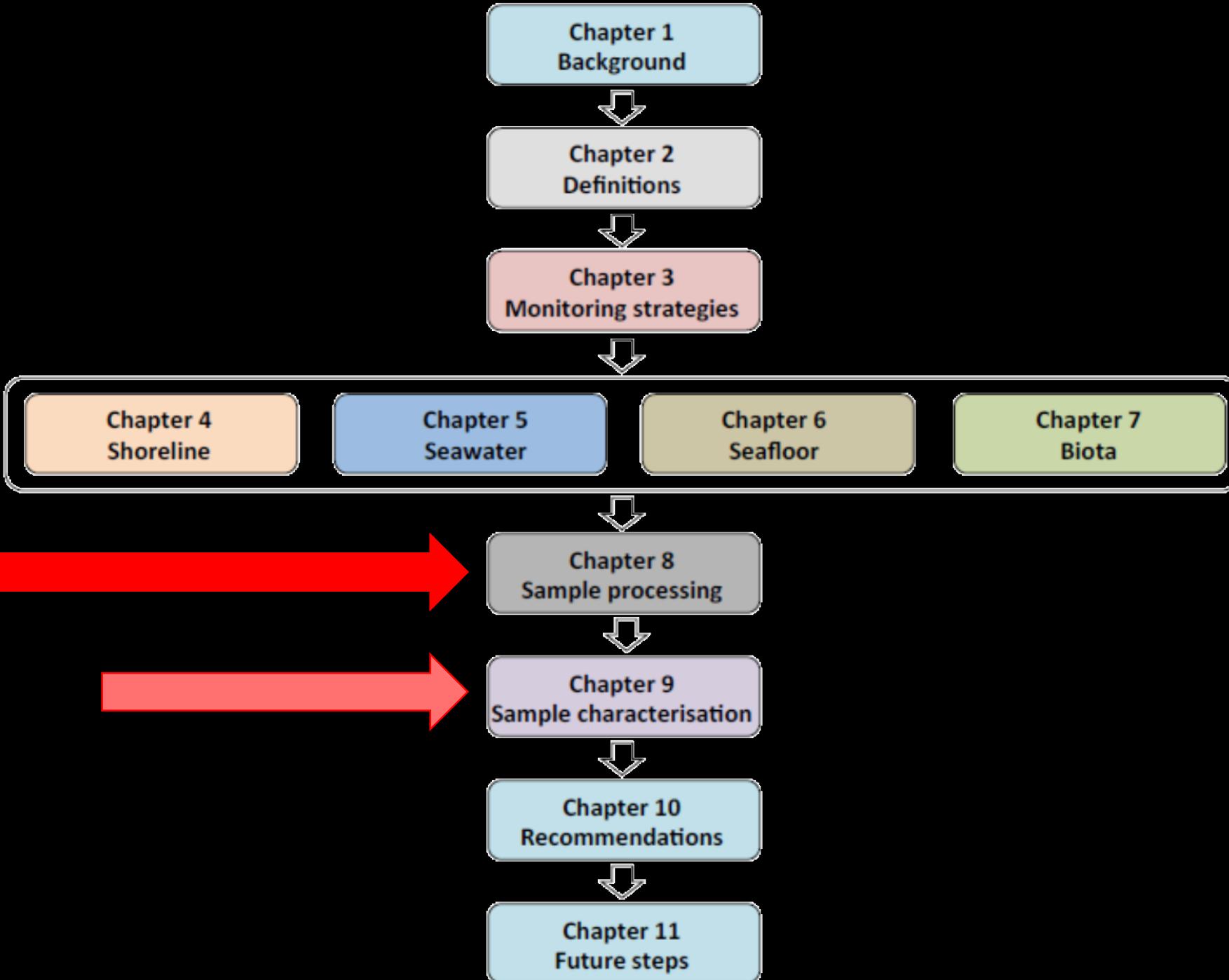
Component	Beach	Seafloor			Seawater		Biota		M
Protocol	Visual	Diving <20m	Trawling <800 m	ROV*	Trawl	Ship surveys	Ingested	Entanglement	M
Sampling	L	M	M	VH	L	M	M/H	M/H	L
Processing	L	L	L	M	L	M	H	M	M

Table 10.5 Overview of sampling protocols for different plastic size categories in two compartments. R = recommended, F = feasible.

Compartment	Size	Recommendation	Method	Comments
Water Surface	Mega	F	Aerial survey	Expensive to charter a plane.
	Mega	R	Visual survey	Use ship as the platform to conduct survey.
	Macro	R	Visual survey	See above
	Meso	R	Net tow	Affordable and feasible
	Micro	R	Net tow	Affordable and feasible
	Micro	F	Bulk water pump	Costs involve microplastic
Water Column	Mega	F	Fisheries observer	Cost effective
	Macro	F	Fisheries observer	Cost effective
	Meso	F	Bulk water pump	Costs involve microplastic
	Meso	R	Underway sampling	Cost effective after training.
	Meso	F	Bongo net	Need vessel
	Micro	F	Bulk water pump	Costs involve microplastic
	Micro	R	Underway sampling	Costs involve microplastic
	Micro	F	Bongo net	Need vessel

Table 10.4. Overview of sampling protocols for different litter size categories at three main shoreline types: Sandy Beaches, Rocky Shores (including cobble and boulder beaches) and Mangroves and Salt Marshes. R = recommended, F = feasible. Values in parentheses indicate approximate transect widths to sample for different litter size categories.

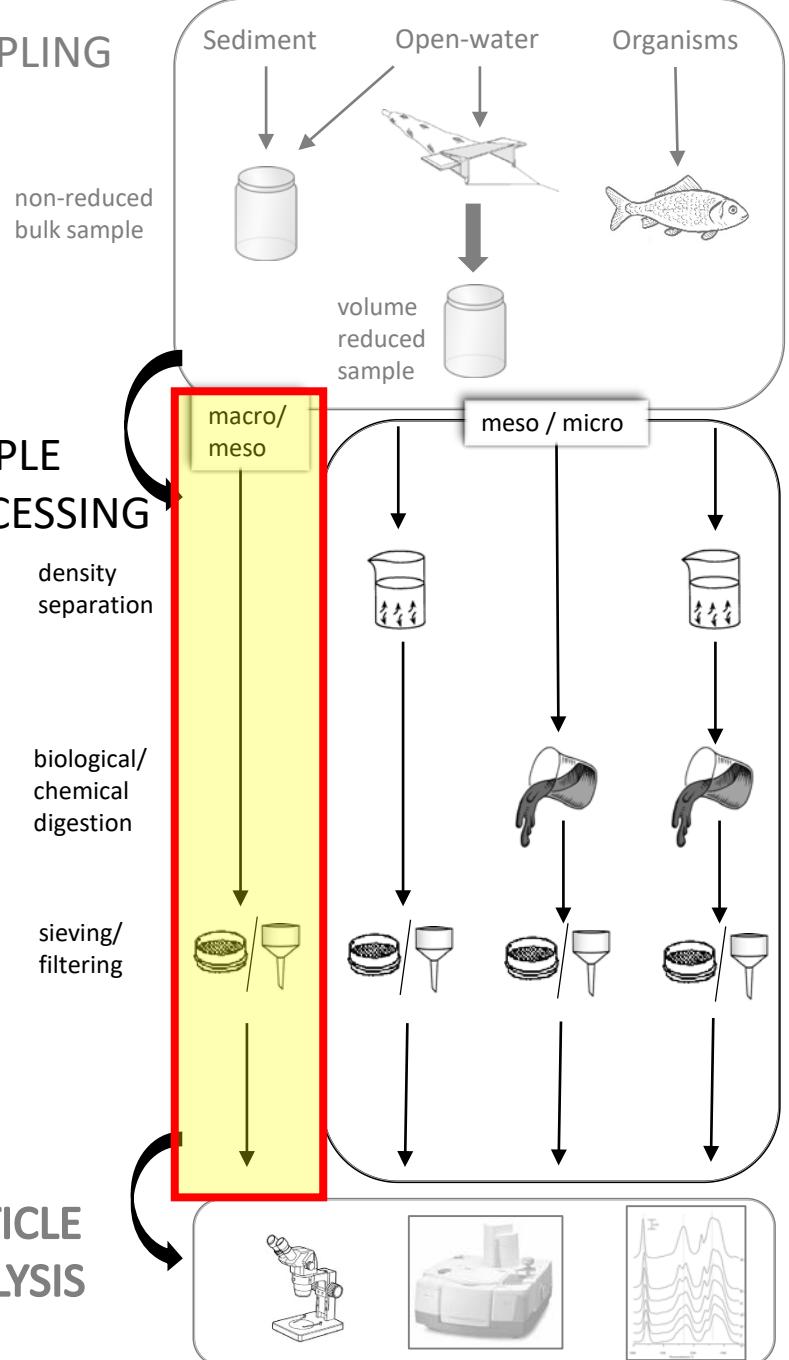
Survey goal	Size	Sandy beaches	Rocky shores	Mangroves and Salt Marshes	Comments			
Baseline surveys	Mega	R	F	F	One-off visual surveys			
	Macro Surface	R	F*	F	One-off visual surveys			
	Macro Buried	F			Sieve to collect litter; sample to at least 10 cm deep			
	Meso	R			Sieve to collect litter to ≥5-10 cm deep			
	Micro	F (cores)*		F (cores)	Surface sieving or sediment cores			
			F		Mark litter and resample at regular intervals			
Survey goal	Size	Marine mammals	Birds	Fish	Invert-brates	Corals	Epibionts	Remarks
Ingestion	Mega	F						Opportunistic, strandings
	Macro	F	R					OSPAR monitoring
	Meso	F	R					
	Micro	F	R	R	R	F	F	
Entanglement	Mega	R	R				F	Opportunistic strandings
	Macro	R	R				F	
	Meso				F	F		
	Micro							
Habitat	Mega					R	R	
	Macro		R			F	R	
	Meso		F			F	R	
	Micro						R	



SAMPLING

SAMPLE PROCESSING

PARTICLE ANALYSIS



Chapters
5-7

This
Chapter

Chapter
9

Density Separation

Salt	Density (g cm ⁻³)	Reference
Sodium Chloride (NaCl)	1.2	Hidalgo-Ruz et al. 2012
Sodium Polytungstate (PST)	1.4	Hidalgo-Ruz et al. 2012
Sodium Iodide (NaI)	1.6	Claessens et al. 2013
Zinc Chloride (ZnCl ₂)	1.7	Imhof et al. 2012
Zinc Chloride (ZnCl ₂)	1.6	Zobkov and Esiukova 2017

Biological/Chemical Digestion

Purification method	Advantages
Biological	Inexpensive
Chemical	Rapid (24 h)
Biological	Effective
Chemical	Minimal damage to most polymers
Biological	Effective
Chemical	Minimal damage to most polymers

Fe (II)

30%
 H_2O_2

Open Water



initial

Salt	Density (g cm ⁻³)
Sodium Chloride (NaCl)	1.2

Eriksen et al., 2013

➤ few fibers

since

Purification method
Oxidative digestion

WPO

Mason et al., 2016a (Lake MI)

Sutton et al., 2016 (SF Bay)

Mason et al., 2016b (WWTP)

Baldwin et al., 2016 (Tributaries)

Sediment



Salt	Density (g cm ⁻³)
Zinc Chloride (ZnCl ₂)	1.6

Zobkov and Esiukova 2017

Purification method
Oxidative digestion

WPO

Lenaker et al.,
in prep

Fish



Purification method
Alkaline digestion

Purification method
Oxidative digestion

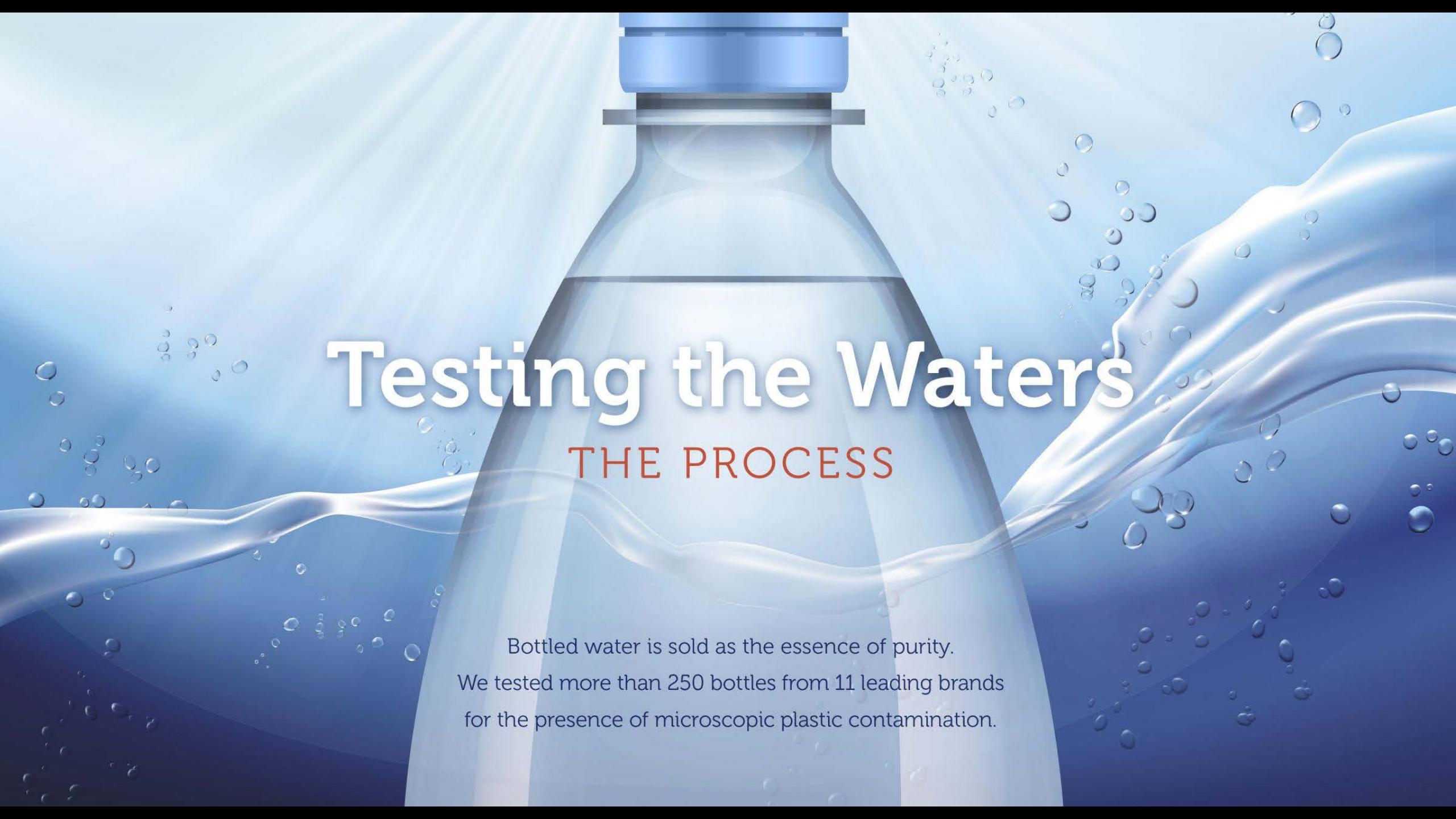
WPO

Ricotta et al.,
unpublished work



BOTTLED WATER

❖ Mason et al., *Frontiers in Chemistry*, 2018.



Testing the Waters

THE PROCESS

Bottled water is sold as the essence of purity.

We tested more than 250 bottles from 11 leading brands
for the presence of microscopic plastic contamination.

01

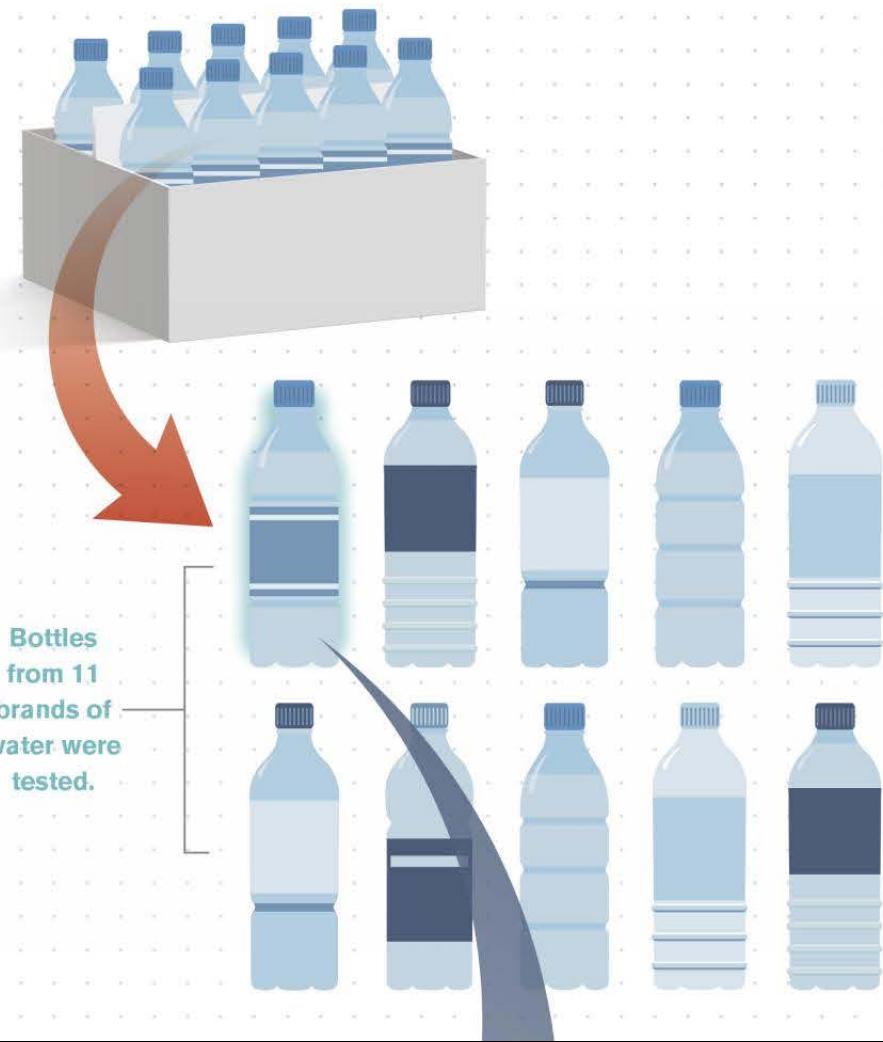
PREPARATION

STAGE.01

FIG. I * The sample

From each case of bottled water, 10 bottles are randomly chosen to be tested.

Where did the bottled water come from? Discover more. 





The bottles are opened in the laboratory under a laminar airflow hood.

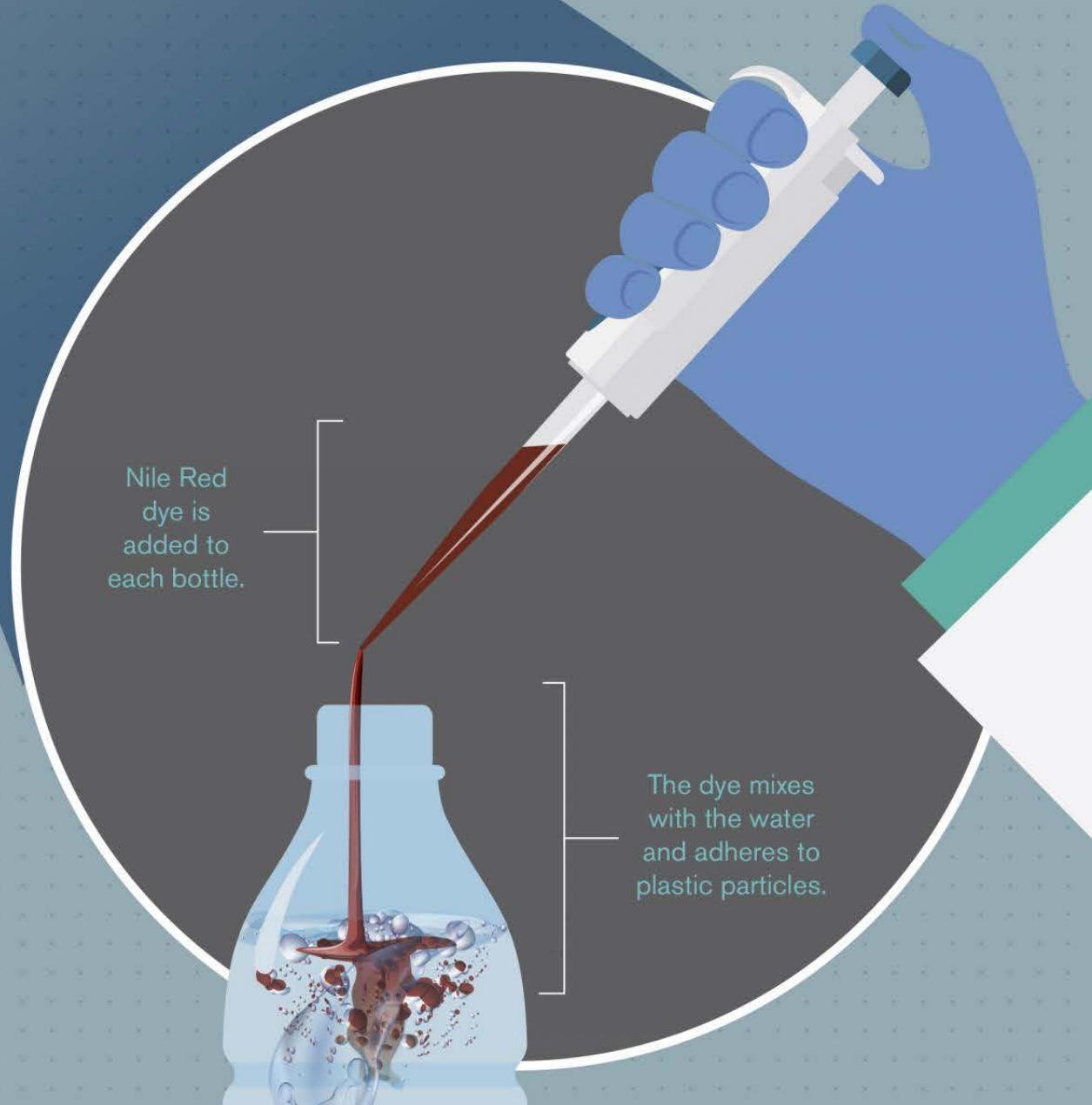
Precautions are taken to avoid contamination.

FIG. II
Adding dye

Each bottle is injected with Nile Red dye, which attaches to plastic particles in the water.



The bottles with the dye stand in the fume hood for 30 minutes.



Nile Red dye adheres to the surface of plastics, but not to naturally occurring materials in the water. The dye fluoresces under blue light.

The dye changes color when diluted in water.

.02

FILTERING

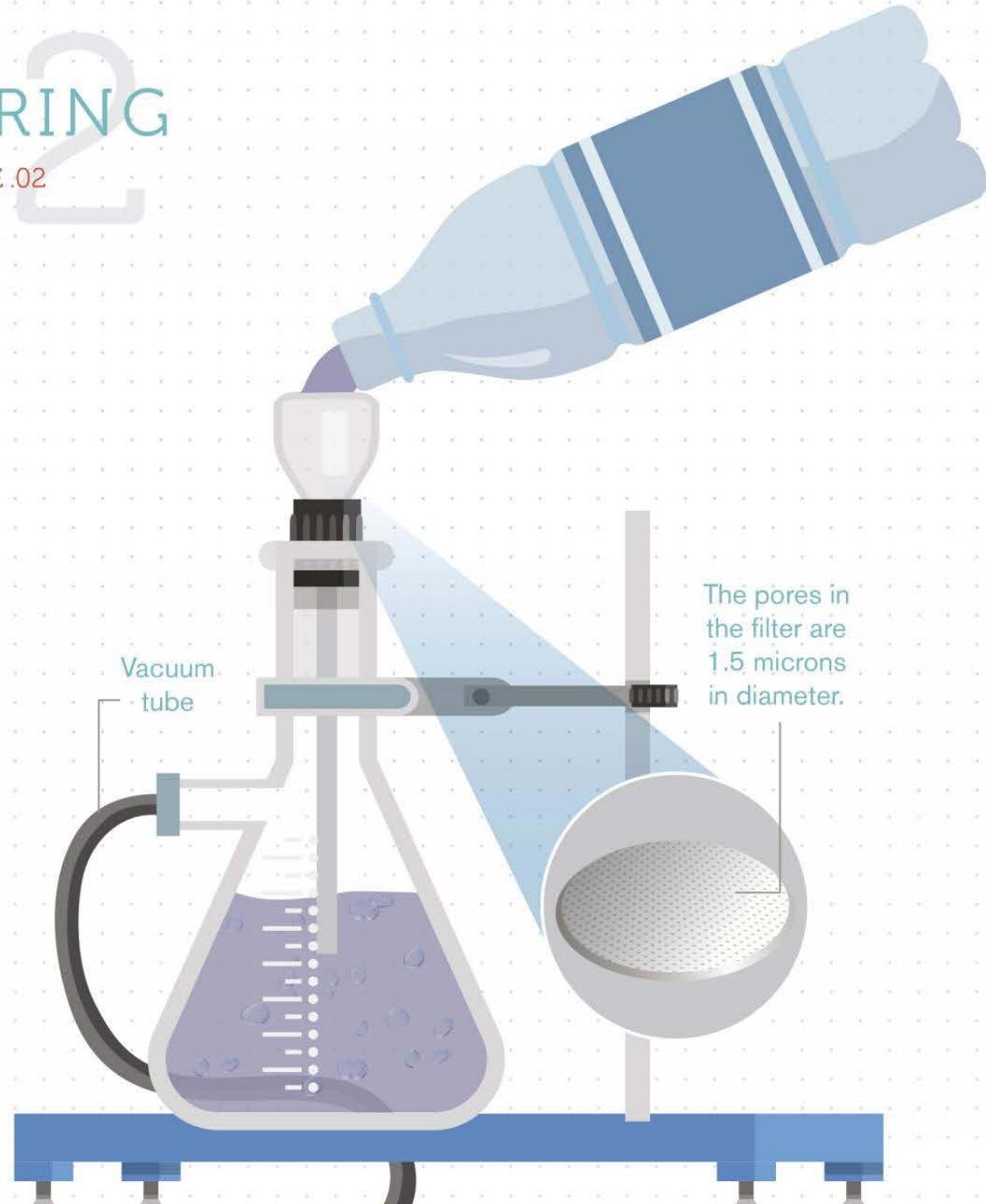
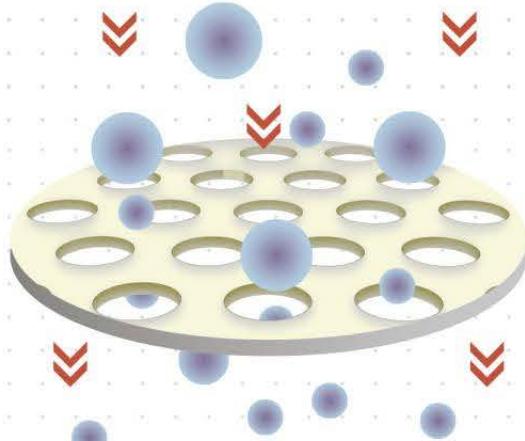
STAGE .02

FIG. III

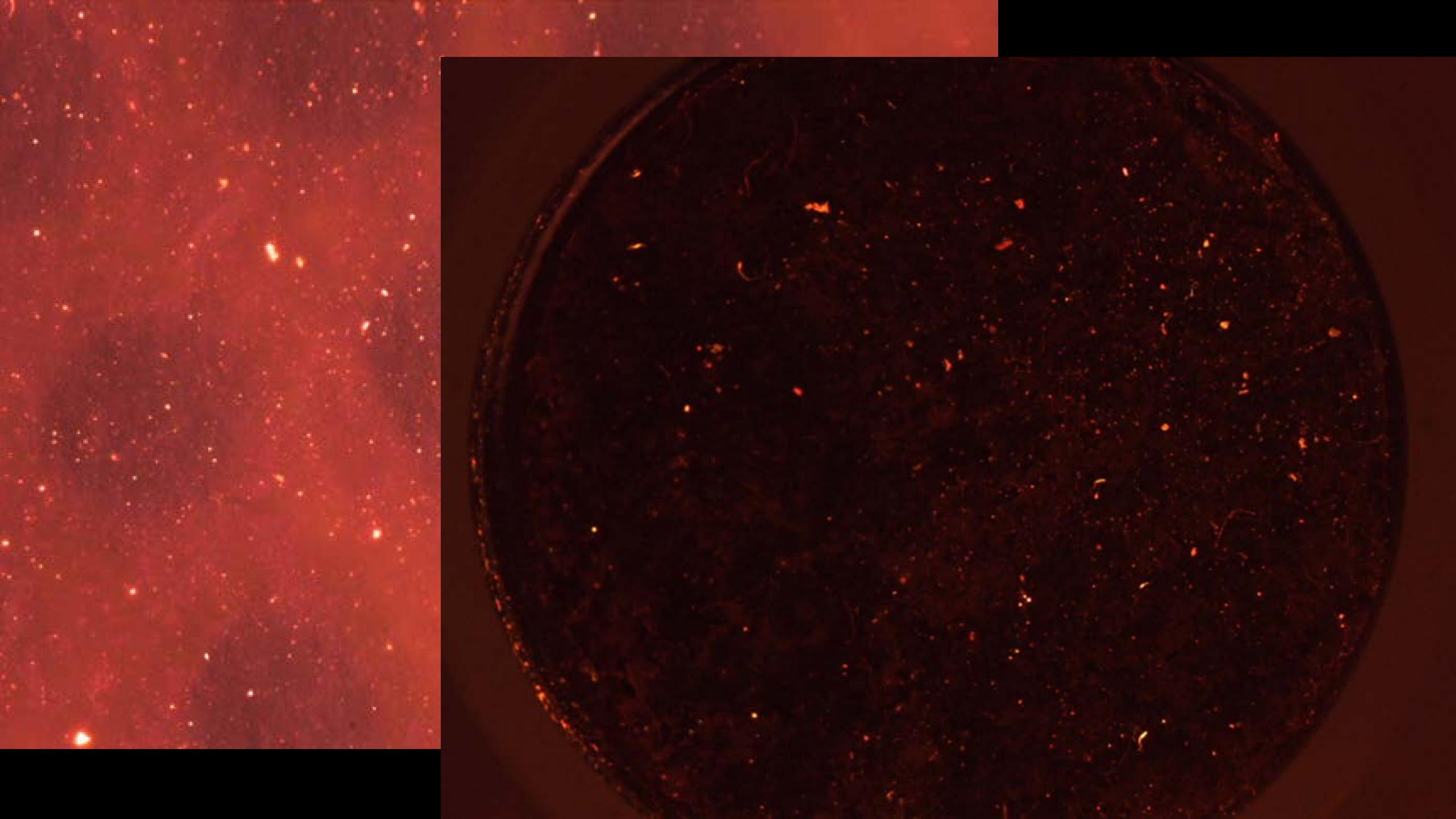
Vacuum filtering

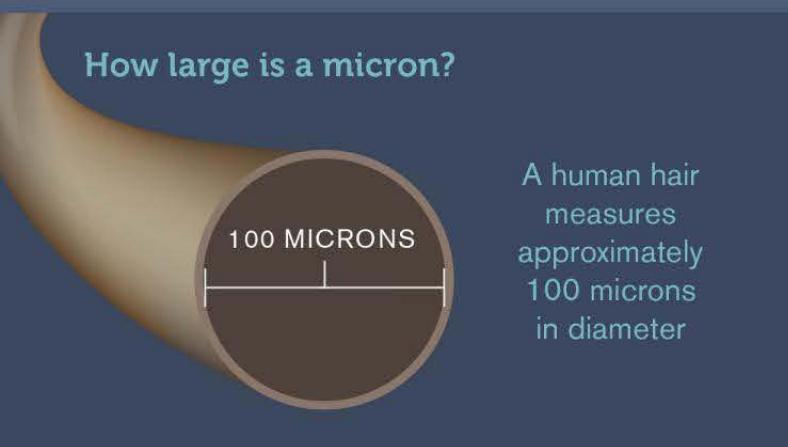
The bottled water with the dye is filtered through glass fiber filters.

Filtering is performed twice to capture particles that may have passed through the first filter.



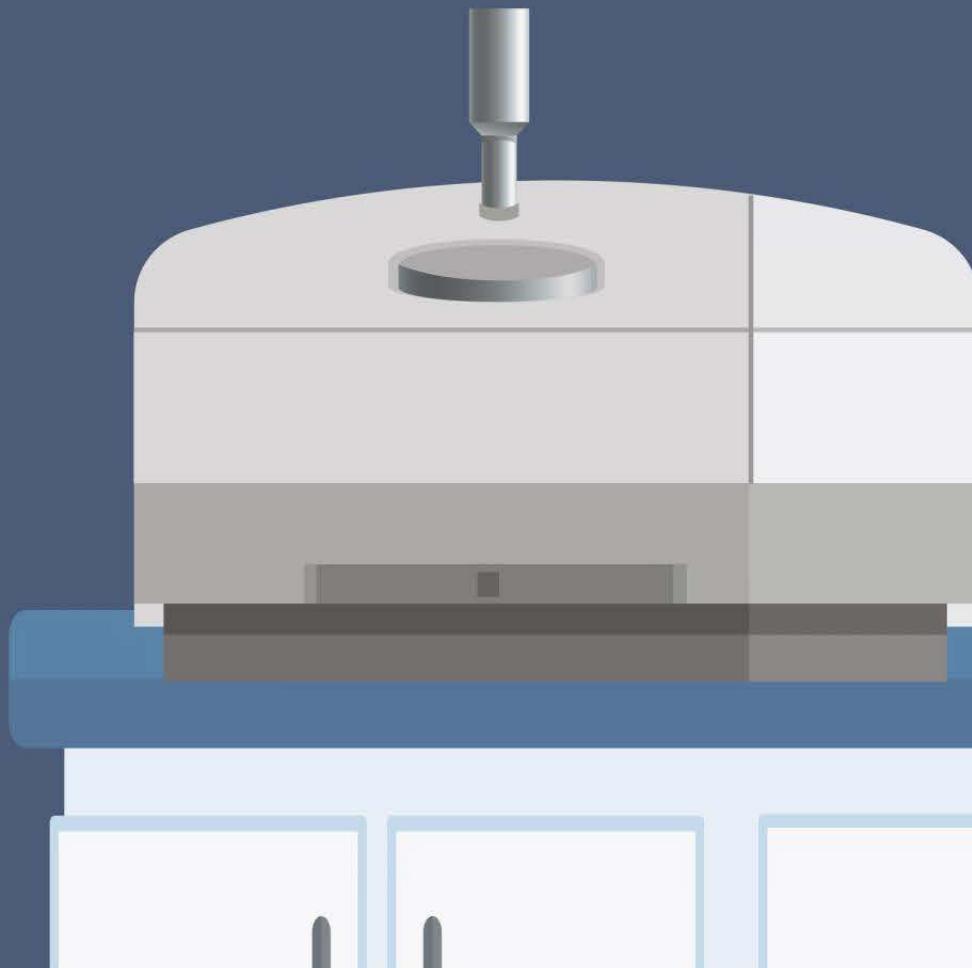






Particles 100 microns or larger are photographed and counted.

The particles are placed in a Fourier-Transform Infrared Spectroscopy (FTIR) machine which uses infrared light to identify the type of plastic.





THE
HEINZ
AWARDS



THE BURNING RIVER
FOUNDATION



New York State



Thank You!



PennState
Behrend

